

### Comparative study of neem leaf (*Azadirachta indica*) suspension and toltrazuril against coccidiosis in Sonali chicken

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#### Abstract

**Background:** Coccidiosis is recognized as the parasitic disease with the greatest economic impact on poultry industries worldwide. Sonali chicken are commonly affected by coccidiosis in Bangladesh. Recently, the concerns about possible drug resistance have aroused great caution in the usage of drug in the animal industry. As an alternative herbal extracts are used and Neem leaves (*Azadirachta indica*) have anti-coccidial property. This study was conducted to evaluate the comparative efficacy of Neem leaves suspension and toltrazuril on coccidial load, blood constituents, growth performance of Sonali chicken infected with *Eimeria tenella*.

**Methods:** The study was conducted in the laboratory and experimental shed under the department of physiology and pharmacology, HSTU, Dinajpur during, 2<sup>nd</sup> January to 3<sup>rd</sup> February, 2019. Eighty (80) Sonali chicken of seven days old were randomly divided into four groups named T<sub>0</sub>, T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> and each group contained 20 birds. All groups were supplied *E. tenella* orally except T<sub>0</sub> group and after 3 days T<sub>2</sub> group was treated with Neem leaves suspension (5%) for 15 days and T<sub>3</sub> groups was treated with toltrazuril (topzuril ®) solution (1 ml/lit) for 2 days.

**Results:** Results showed that protozoal load decreased significantly ( $P < 0.01$ ) in T<sub>2</sub> & T<sub>3</sub> groups whereas increased significantly ( $P < 0.01$ ) in T<sub>0</sub> and T<sub>1</sub> group. The total erythrocyte count and hemoglobin in different treatment groups were similar and the differences were non-significant ( $P > 0.05$ ) statistically except total leukocyte count (TLC). The body weight of T<sub>0</sub>, T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> group at day 30 were 432.65, 469.81, 548.81, 489.22 gm respectively which is statistically significant ( $P < 0.01$ ) and highest body weight gain was recorded from Neem leaves suspension treated group.

**Conclusion:** Neem leaves suspension have significant effects on protozoal load, body weight and no significant effect on haematological parameters except TLC.

**Keywords:** Coccidiosis; neem leaf; toltrazuril; Sonali chicken.

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### Introduction

Bangladesh is an agriculture based tropical country where more than 80% of the country's 160 million people are living in rural areas and highly dependent on agriculture. Poultry farming is one of the major activities in rural areas as it provides immense employment opportunities to the local communities especially for youth & women that helps in poverty reduction, ensures food security and improves the nutritional status of the people. Poultry industry can produce very high quality proteins for human nutrition as well as a source of income for the community in many countries, therefore poultry production plays very important role in economic development of any country (Tarhyel *et al.*, 2012). Poultry is a promising sector in Bangladesh which is increasing day by day. Poultry production is hindered by many problems among which various diseases namely bacterial, viral and parasitic infections are the most important (Ojok, 1993).

The northern part of Bangladesh is considered as a poultry hub especially of Sonali chicken, which is used as meat purpose. Sonali chickens are well adapted in the environmental conditions of Bangladesh, as it requires less care and attention as compared to other breeds. It is also easier for women to rear (Saleque and Saha, 2013). In spite of many positive reasons behind rearing of Sonali chickens, there are some causes that hamper in optimum production of Sonali chicken by smallholder households in Bangladesh (Biswas *et al.* 2006). Poultry diseases are one of the major constraints for developing poultry industry in Bangladesh (Islam and Samad, 2004).

Avian coccidiosis is a parasitic disease of intestinal tract caused by single cell protozoan parasite belonging to genus *Eimeria*. It causes massive destruction of the epithelial cells, which leads to bloody diarrhea, reduced weight gain and temporary reduction in egg production (Dalloul and Lillehoj, 2005; Razzaq *et al.*, 2011).

Coccidiosis has been considered as a very harmful disease affecting growth and performance of poultry (Lin *et al.*, 2006; Mujahid *et al.*, 2007; Bachaya *et al.*, 2012). Thus coccidiosis is probably the most expensive and

wide spread infectious disease in commercial poultry systems.

In Bangladesh, a number of drugs are available for the treatment and prevention of chicken coccidiosis. Among these sulphaclozine sodium are used extensively throughout the country. In addition, ionophorous antibiotic i.e., maduramycin and lasalocid and some other drugs like amprolium, toltrazuril are also used in the field. It has been found that the efficacy of various anti-cocccial drugs varies considerably against coccidiosis (Lee, 1978).

Toltrazuril is a symmetrical triazinetrione compound and 2.5% oral solution has been shown to be effective against all species of *Eimeria* infecting chickens (Mehlhorn *et al.*, 1988). It is active against all intracellular developmental stages.

Chemoprophylaxis with Toltrazuril enhances immunity against poultry coccidiosis (Chapman, 1987). It has been proved that therapeutic medication with Toltrazuril protects the birds from clinical coccidiosis (Ramadan *et al.*, 1997).

Drug resistance is very much alarming matter nowadays. New drug resistance mechanisms are emerging and spreading globally, threatening our ability to treat common infectious diseases, resulting in prolonged illness, disability, and death. So, we need to quickly replace of anticoccidial with other options. These new options should be inexpensive and available in everywhere as well as healthy for human society. It is thought that medicinal plants are a good alternative for this purpose. Since ancient times medicinal plants play important role in health management of traditional poultry production (Eevuri and Putturu, 2013). In recent years, the growing demand for herbal product has led to a quantum jump in volume of plant materials traded within and across the countries.

Secondary plant metabolites (Phytochemicals), previously with unknown pharmacological activities, have been extensively investigated as a source of medicinal agents (Krishnaraju *et al.*, 2005). Thus it is anticipated that phytochemicals with adequate anticoccidial efficacy will be used

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for the treatment of the protozoal infections. Also, in contrast to anticoccidial, most active components of medicinal plants are readily absorbed in intestine and have short half life.

Neem elaborates a vast array of biologically active compounds that are chemically diverse and structurally complex. More than 140 compounds have been isolated from different parts of Neem.

The tender leaves along with piper nigrum Linn are found to be effective in intestinal helminthiasis. Since Neem leaf increases immunity against some common diseases which assists to reduce mortality, it is likely that the growth pattern of sonali chicks will proceed uninterrupted.

So, this study was undertaken to evaluate the comparative efficacy of Neem leaves suspension and toltrazuril on coccidial load, blood constituents, growth performance of sonali chicken infected with *Eimeria tenella*.

#### Materials and Method

The experiment was conducted at the experimental shed under the department of Physiology and Pharmacology, for a period of 65 days from 2<sup>nd</sup> January to 3<sup>rd</sup> February, 2018.

#### Preparation of house

First the room as well as the wire cages were washed by sweeping with tap water using hose pipe connected with the tap. The room was properly ventilated and disinfected with a phenolic disinfectant and allowed to dry leaving the room unused with the electric fan and the bulb switched on overnight. All the utensils required for the experiment such as feeder, water pot, beakers, pestle and mortar, syringe, needle etc. were set in the experimental shed.

#### Grouping of birds

Eighty (80) Sonali chicken were used to evaluate the dietary effect of Neem leaf suspension on *Eimeria tenella* infection, growth performance. The Sonali chicken were assigned into four groups containing 20 birds in each groups. 20 birds kept in each group were considered as an

experimental unit. Sonali chicken were randomly distributed in every groups.

**Group T<sub>0</sub>:** The Sonali were fed normal diet and given water ad-libitum and their body weight was recorded at every 15 days interval. Body weights, blood parameter and *Eimeria tenella* protozoal load were measured at the times when that of other groups were measured. This group was served as “Negative control” group.

**Group T<sub>1</sub>:** The Sonali chicken were supplied with protozoa after acclimatization to induce *Eimeria tenella* infection in this group. No antiprotozoal treatment was given against *Eimeria tenella* in T<sub>1</sub> group. Adequate feed and drinking water was given. This group served as “Positive control” group.

**Group T<sub>2</sub>:** The Sonali chicken were supplied with *Eimeria tenella* protozoa after acclimatization to induce *Eimeria tenella* infection as like as T<sub>1</sub> group. This group was left untreated for 24 hours to establish *Eimeria tenella* infection. After 24 hours, this group was treated with Neem leaves suspension as herbal antiprotozoal agent at a dose rate of 0.25 ml per 100 ml of drinking water. This group served as “Neem leaf suspension” group.

**Group T<sub>3</sub>:** After acclimatization Sonali chicken of this group were supplied with *Eimeria tenella* protozoa as like as T<sub>1</sub> & T<sub>2</sub> group. This group was left untreated for 24 hours to establish *Eimeria tenella* infection. After 24 hours, this group was treated with antiprotozoal drug (Topzuril liquid @ 1 ml per 2lit water). This group served as “Topzuril liquid” group.

#### Collection of feed

Polli mash commercial feed were collected from Griholokkhi Poultry Feed, Kalitola, Dinajpur from a reputed Sonali feed exporter. Mash and water were provided *ad-libitum* during the whole experimental period.

The ingredients that were used for the composition of mash feed are described in Table 1.

Table 1. Feed formulation

Name of ingredients	Percentage of ingredients	Required Amount of ingredients (kg)	Rate of ingredients/kg (TK)	Cost of ration (TK)
Maize	59	4.956	23	113.988
Soybean meal	22	1.848	58	107.184
Protein conc.	5	0.42	80	33.60
Rice polish	3	0.252	20	5.04
Saw dust	8	0.672	05	3.36
Oil	2	0.168	110	18.48
Lime stone	1	0.084	08	0.672
Enzyme	Trace amount			
Total	100	8.4		282.324

Metabolizable Energy (Kcal/100kg) was 3009.11 and Crude Protein was 18.95%  
Feed cost per kg 33.61 Taka

### Acclimatization of Sonali chicken

Immediately after reaching the destination the sonali were shifted to wire cages. They were fed with Griholokkhi Poultry Feed, Kalitola, Dinajpur and drinking water *ad-libitum*. Glucose and vitamin C were supplied with drinking water for first three days to overcome the transportation stress. Sonalis were allowed to acclimatize in their new environment for 13 days before the commencement of the experiment.

### Lighting

During the whole experimental period, all Sonali chicken were exposed to a 16 hours continuous photoperiod (natural light plus artificial light) in an open sided house. Electrical bulbs were used for additional light at night.

### Collection and processing of neem leaves

Neem leaves were collected from the HSTU, Dinajpur. Young Neem leaves were collected and washed with fresh water. Before chopping it into small pieces, it was soaked with cotton for removing the adhesive water. Then the leaf was chopped into small pieces and was mashed with the help of pestle and mortar. Leaf juice was collected by squeezing mashed leaf. Then it had been produced 0.25% of suspension of grinded Neem leaf with distilled water.

### Collection, preservation and administration of toltrazuril

Topzuril liquid was collected from Square pharmaceuticals Ltd. It was preserved in a dry place at room temperature and was administered

at a dose rate of 1ml per 2 litre of drinking water for 3-7 days.

### Recording of body weight

The effect of the Neem leaves extract and toltrazuril on body weight was recorded before and during administration of drugs.

Chickens under trial and control groups were weighed with Electric weighing machine. The weight of each chicken was taken before feeding in the morning, in noon and afternoon. The average of these three weights was calculated and recorded.

### Collection and preparation of test organism

The test organism (*Eimeria tenella*) was collected from the Gopalgonj local farm area, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur.

### Collection and examination of feces for parasitic oocyst investigation

Feces were examined by two different types of qualitative tests; namely direct smear and flotation techniques were used to examine the fecal samples to identify the morphological features of oocysts (Hendrin and Robinson, 2006; Soulsby, 1982).

### Hematological parameters

Blood samples were collected from the wing vein of chicken of both control and treated groups at pre-feeding and during feeding (28 days) period at 7 days interval to study the effect of the Neem

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leaves extract and Toltrazuril. The following parameters were observed:

- Total Erythrocyte Count (TEC)
- Hemoglobin Estimation (Hb)
- Packed Cell Volume (PCV)
- Erythrocyte Sedimentation Rate (ESR)
- Total Leukocyte Count (TLC)

Total Erythrocytes Count (TEC), Erythrocytes Sedimentation Rate (ESR), Packed Cell Volume (PCV) and Total Leukocyte Count (TLC) were performed as per methods described by Schalm (1967). Hemoglobin estimation was performed as per method described by Coffin (1955).

#### Statistical analysis

The data were analyzed statistically between control and treated groups of chicken by using SPSS version 22 and Microsoft Excel. Statistically significant differences between group means were determined by analysis of variance (ANOVA).

### Results and Discussion

#### Anticoccidial effect of Neem leaves suspension and toltrazuril liquid

Coccidial load in feces of *E. tenella* infected Sonali chicken were presented in Table 1. This study showed that, coccidial load in feces varied significantly among the treated groups. Coccidial load significantly ( $P < 0.01$ ) increased in  $T_0$  (negative control) and  $T_1$  (positive control) group. Coccidial load significantly ( $P < 0.01$ ) decreased in  $T_2$  group supplied with 1% Neem leaves suspension and toltrazuril liquid suspension. Karim *et al.* (2016) also observed similar observation.

#### Total Erythrocyte Count

Total Erythrocyte Count (TEC) is presented in Table 2. The values of TEC in all treated groups and control group were similar and within the normal range at 15th and 30th day of experimental period.

Table 2. Estimation of oocyst per gram (OPG) during experimental period

Days	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	Significant level
1 day	0.00±0.00 <sup>a</sup>	319.31±1.67 <sup>b</sup>	412.48±0.92 <sup>c</sup>	431.48±1.25 <sup>d</sup>	*
15 day	0.00±0.00 <sup>a</sup>	390.89±1.81 <sup>c</sup>	190.36±1.56 <sup>b</sup>	0.00±0.00 <sup>a</sup>	*
30 day	0.00±0.00 <sup>a</sup>	387.28±1.62 <sup>c</sup>	55.68±1.09 <sup>b</sup>	0.00±0.00 <sup>a</sup>	*

Values are expressed as mean ± standard error of means.

a, b, c Means in each row with different superscript are significantly different at (0.05).

\* = Significant at 1% level of significance.

T<sub>0</sub> = Control (Basal diet),

T<sub>1</sub> = (Basal diet plus *E. tenella*),

T<sub>2</sub> = (Basal diet plus *E. tenella* plus 0.5% neem leaves suspension),

T<sub>3</sub> = (Basal diet plus *E. tenella* plus toltrazuril @ 1 ml per 1 litre drinking water)

Table 3. TEC in different groups of Sonali chicken

Days	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	Significant level
15 day	2.97±.18	3.19±.21	3.14±.16	3.28±.06	NS
30 day	3.15±.22	3.47±.09	3.35±.07	3.34±.06	NS

NS= Non significant

Values are expressed as mean ± standard error of means. Means within column and between column are not statistically significant ( $P > 0.05$ ). NS=Not significant.

#### Total Leukocyte count (TLC)

The values of TLC are presented in Table 4. The values of TLC of T<sub>0</sub> (Basal diet) and T<sub>3</sub> (*E. tenella* plus toltrazuril liquid) group were similar. But in case of T<sub>1</sub> (Basal diet plus *E. tenella*) and T<sub>2</sub> (*E. tenella* plus Neem leaves suspension) groups the values differed.

#### Hemoglobin

Hemoglobin (Hb) is presented in Table 5. The values in all treated groups and control group were similar and the values were within normal range.

Table 4. TLC in different groups of Sonali chicken

Days	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	Significant level
15 day	6.39±.05 <sup>b</sup>	6.27±.05 <sup>d</sup>	6.83±.14 <sup>c</sup>	6.58±.21 <sup>a</sup>	*
30 day	6.35±.07 <sup>d</sup>	6.09±.28 <sup>b</sup>	6.75±.12 <sup>a</sup>	6.66±.07 <sup>c</sup>	*

Values are expressed as mean ± standard error of means.

a, b, c Means in each row with different superscript are significantly different at (0.05).

\* = Significant at 1% level of significance.

T<sub>0</sub> = Control (Basal diet),

T<sub>1</sub> = (Basal diet plus *E. tenella*),

T<sub>2</sub> = (Basal diet plus *E. tenella*. plus 0.5% neem leaves suspension),

T<sub>3</sub> = (Basal diet plus *E. tenella*. plus toltrazuril @ 1 ml per 1 litre drinking water)

Table 5. Estimation of Hemoglobin during experimental period

Days	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	Significant level
15 day	7.67±.44	7.54±.16	7.90±.02	8.04±.06	NS
30 day	8.05±.05	7.53±.25	8.25±.07	8.41±.03	NS

NS= Non significant

Values are expressed as mean ± standard error of means. Means within column and between column are not statistically significant (P>0.05). NS=Not significant.

### Body Weight

Table 6 indicated that there were a significant (P<0.01) increase in body weight in T<sub>2</sub> (Neem leaf suspension) and T<sub>3</sub> (topzuril liquid) group than the T<sub>0</sub> (Basal diet) and T<sub>1</sub> (*E. tenella*). Birds treated with Neem leaves suspension (T<sub>2</sub> group)

had the highest body weight gain compared to other groups. Lakkundi *et al.* (2002) also observed the same effect of toltrazuril and amprolium on body weight and feed efficiency of broiler chicken experimentally infected with *Eimeria tenella*.

Table 6. Estimation of Body Weight during experimental period

Days	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	Significant level
1 day	180.84±.51	182.01±0.93	152.43±0.94	182.41±0.87	NS
7 day	223.47±.05 <sup>d</sup>	210.54±1.43 <sup>a</sup>	250.31±0.71 <sup>b</sup>	261.92±1.42 <sup>c</sup>	*
14 day	253.50±1.18 <sup>b</sup>	264.23±0.84 <sup>c</sup>	343.72±1.55 <sup>d</sup>	314.11±0.43 <sup>a</sup>	*
21 day	375.70±0.4 <sup>d</sup>	351.33±1.49 <sup>c</sup>	497.37±1.89 <sup>b</sup>	362.42±1.64 <sup>a</sup>	*
30 day	432.65±0.71 <sup>b</sup>	469.81±1.23 <sup>c</sup>	548.86±1.29 <sup>d</sup>	489.22±1.33 <sup>a</sup>	*

Values are expressed as mean ± standard error of means.

a, b, c Means in each row with different superscript are significantly different at (0.05).

\* = Significant at 1% level of significance,

T<sub>0</sub> = Control (Basal diet) ,

T<sub>1</sub> = (Basal diet plus *E. tenella*),

T<sub>2</sub> = (Basal diet plus *E. tenella*. plus 0.5% neem leaves suspension),

T<sub>3</sub> = (Basal diet plus *E. tenella*. plus toltrazuril @ 1 ml per 1 litre drinking water)

### Conclusions

Nneem leaves (*Azadirachta indica*) suspension and toltrazural have anticoccidial effect and effects on growth performance of sonali chicken with *E. tenella*. It can be used for the control of coccidiosis and as a good growth promoter, leukocyte enhancer in Sonali chicken. It may be stated that the neem leaf (*Azadirachta indica*) may provide a new therapeutic avenue against *E.*

*tenella* because of its availability in our country and effectiveness.

### Conflict of interest

None to declare.

### References

1. AL-Fifi ZIA. Effect of leaves extract of *Carica papaya*, *Vernonia amigdalina* and *Azadirachta indica* on coccidiosis in free-range chickens.

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- Asian Journal Animal Science. 2007;1(1): 26-32.
2. Beraa AK, Bhattacharyaa D, Pana D, Dharab A, Kumar S, Das SK. Evaluation of Economic Losses due to Coccidiosis in Poultry Industry: Agricultural Economics Research Review. 2013; 22: 91-96.
  3. Bhajoni PS, Meshram GG, Lahkar M. Evaluation of the antiulcer activity of the leaves of *Azadirachta indica*: An experimental study. Integrative Medicine International. 2016; 3(1-2): 10-16.
  4. Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. Biological activities and medicinal properties of Neem (*Azadirachta indica*). Current Science. 2002; 82: 1336-1345.
  5. Bais B, Purohit GR, Dhuria RK, Pannu U. Nutritive value of sares and Neem leaves in marwari goats. Indian Journal of Animal Nutrition. 2002; 19(3): 266-268.
  6. Biu, AA, Yusuf SD, Rabo JS . Use of Neem (*Azadirachta indica*) aqueous extract as a treatment for poultry coccidiosis in Borno, Maiduguri, Nigeria. African Scientist. 2006; 7:3.
  7. Biswas PK, Uddin GM, Barua H, Roy K, Biswas D, Ahad A, Debnath NC . Causes of loss of Sonali chickens on smallholder households in Bangladesh. Preventive Veterinary Medicine. 2006; 76(3-4): 185-195.
  8. Bhowmik D, Chiranjib, YJ, Tripathi KK, Kumar KS. Herbal remedies of *Azadirachta indica* and its medicinal application. Journal of Chemical and Pharmaceutical Research. 2010; 2(1): 62-72.
  9. Bujmehrani H. Prevalence and risk factors for subclinical coccidiosis in broiler chicken farms in Mazandaran province, Iran. Tropical Animal Health and Production. 2011; 43: 16011604.
  10. Chapman HD. In: Avian coccidiosis research: Poultry Science. 2014; 93 (3): 501511
  11. Chapman HD, Barta JR, Blake D, Gruber A, Jenkins M, Smith NC, Suo X, Tomley FM. A selective review of advances in coccidiosis research. Advances in Parasitology. 2013; 83: 93-171.
  12. Yun CH, Lillehoj HS, and ELillehoj EP. Intestinalimmune responses to coccidiosis. Developmental and Comparative Immunology. 2000; 24 (2-3): 303–324.
  13. Dhillon NK., Jasmer S, Bal MS and Ashuma T. Efficacy of toltrazuril against different levels of *Eimeria tenella* infection in chicken. Journal of Research, Punjab Agricultural University. 2004; 41(1): 150-155.
  14. Dalloul RA and HS Lillehoj. Recent advances in immunomodulation and vaccination strategies against coccidiosis. Avian Disease. 2005; 49: 1-8.
  15. Dash SP. Phytochemical and Biochemical Characterizations from Leaf Extracts from *Azadirachta indica* : An Important Medicinal Plant. Biochemistry Analytical Biochemistry. 2017; 6:2. DOI: 10.4172/2161-1009.1000323.
  16. Dholi SK, Raparla R and Mankala SK. Invivo Antidiabetic evaluation of Neem leaf extract in alloxan induced rats. Journal of Applied Pharmaceutical Science. 2011; 01 (04): 100-105.
  17. Eevuri T. R and Putturu R. Use of certain herbal preparations in broiler feeds -A review. Veterinary World. 2013; 6(3): 172-179.
  18. Gabriel, S. Mallet, M. Leconte, G. Fort, and M. Naciri. Effects of whole neem leaves feeding on the development of coccidial infection in Sonali chickens. Poultry Science. 2003;82 (11): 1668–1676.
  19. Greuel E and Ruhrmann U. Effectiveness of various dosage concentration and administration times of Baycox medication in experimental chicken coccidiosis in cage conditions. Dtsch Tierarztl Wochenschr. 1986; 93: 29-33.
  20. Grief G. Immunity to coccidiosis after treatment with toltrazuril. Parasitology Research. 2000; 86 (10): 787-790.
  21. Ghanem MM, Radwaan ME, Moustafa AMM, Ebeid MH. Comparative therapeutic effect of toltrazuril, sulphadimidine and amprolium on *Eimeria bovis* and *Eimeria zuernii* given at different times following infection in buffalo calves (*Bubalus bubalis*). Preventive Veterinary Medicine. 2008; 84: 161-170
  22. Gjerde B, Helle O. Chemoprophylaxis of coccidiosis in lambs with a single dose of toltrazuril. Veterinary Parasitology. 1991; 38(2-3):97-107.
  23. Ghazala N, Qazi AW, Gill, ZJ, Rizwan Q, Sajid MA. Evaluation of 2.5 percent Toltrazuril against coccidiosis in broiler chicks. Pakistan

- Journal of Veterinary Research. 2003; 1(2): 8-11
24. Grilli G, Giussani N, Ceruti R, Gavazzi L, Pisoni AM, Ferrazzi V, Gallazzi D. Use of toltrazuril treatment for the control of broiler coccidiosis. *Large Animals Review*. 2003; 9(6): 123-124.
  25. Harder A, Haberkorn A. Possible mode of action of toltrazuril: studies on two *Eimeria* species and mammalian and *Ascaris suum* enzymes. *Parasitology Research*. 1989; 76 (1): 8-12.
  26. Hadas G, Mebrhatu G, Abebe T. Prevalence of Poultry Coccidiosis in Gondar Town, North West Ethiopia. *American-Eurasian Journal of Scientific Research*. 2014; 9 (5): 129-135.
  27. Islam MT, Samad MA. Mortality in chicks associated with economic impact and prospect of layer chick rearer package programme of the participatory livestock development project in Bangladesh. *International Journal of Poultry Science*. 2004; 3(2): 119-123.
  28. Islam MK, Mondal MMH, Quadir ANMA. Prevalence and distribution of coccidiosis in dead chicken. *The Bangladesh Veterinarian*. 1996; 13(12): 32-33.
  29. Joyner L, Long P. Specific characters of the *Eimeria*, with special reference to the *Coccidia* of the Avian Pathology. 2008; 3 (3): 145-157.
  30. Kotsch M, Alnassan AA, Shehata AA. Efficacy of early treatment with toltrazuril in prevention of coccidiosis and necrotic enteritis in chickens. *Avian Pathology*. 2013; 42 (5): 482-490.
  31. Khan MN. Anticoccidial activity of herbal complex in broiler chickens challenged with *Eimeria tenella*. *Parasitology*. 2012; 139(2):237-43.
  32. Karim MA, Lovelu MA, Talukder MH, Alam MZ. Evaluation of anti-coccidial drug sensitivity against experimental coccidiosis in broiler chicks. *Bangladesh Agricultural University*. 2016; 14(1): 57-61.
  33. Krishnaraju AV, Rao TVN, Sundararaju. Assessment of bioactivity of Indian medicinal plants using Brine shrimp (*Altenaria salania*) lethality assay. *International Journal of Applied Science and Engineering*. 2005; 2: 125-134.
  34. Karim MJ. Intra specific variations in avian coccidia of British and Bangladeshi origin. Ph.D. Thesis submitted to the University of Liverpool, England, August;1988, p. 34.
  35. Karim MJ, Begum N. Morphological and biological characterization of chicken *Eimeria* with special reference to species identification. *Veterinary Review*. 1994; 9(1): 7-9.
  36. Karim MJ, Begum N, Khan MSR. Prevalence of coccidiosis in broilers on two farms in Bangladesh. *Pakistan Journal of Scientific and Industrial Research*. 1994; 37(8): 339-341.
  37. Lakkundi JN, Jagannath MS, D'Souza PE. Effect of toltrazuril and amprolium on body weight and feed efficiency of broiler chicken experimentally infected with *Eimeria tenella*. *Indian Journal of Animal Sciences*. 2002; 72(10): 835-837.
  38. Lin H, Decuypere E, Buyse J. Acute heat stress induces oxidative stress in broiler chickens. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*. 2006; 144: 11-17.
  39. Meskerem A, Chaiwat B, Nirat G, Montakan V. Hematological, Biochemical and Histopathological Changes Caused by Coccidiosis in Chickens. *Kasetsart Journal (Nat. Sci.)*. 2013; 47: 238-246.
  40. Michalczyk M, Raś-Noryńska M, Sokół R. Efficacy of toltrazuril (Baycox) for *Eimeria* spp. control in racing pigeons. *Medycyna Weterynaryjna*. 2011, 67: 406-408.
  41. Mathis GF, Froyman R, Kennedy T. Coccidiosis control by administering toltrazuril in the drinking water for a 2-day period. *Veterinary Parasitology*. 2004, 121: 1-9.
  42. Mazurkiewicz M, Podlewska D, Wachnik Z. Coccidiosis in Japanese quails. *Medycyna Weterynaryjna*. 1967; 23: 536-537.
  43. Mehlhorn H, Schmahl G, Haberkorn A. Toltrazuril effective against a broad spectrum of protozoan parasites. *Parasitology Research*. 1988; 75 (1): 64-66.
  44. N. Shiotani E, Baba, Fukata T, Arakawa A, Nakanishi T. Distribution of oocysts, sporocysts and sporozoites of *Eimeria tenella* and *Eimeria maxima* in the digestive tract of chicken. *Veterinary Parasitology*. 1992; 41(1-2):17-22.
  45. Ojok L. Diseases as important factor affecting increased poultry production in Uganda. *Der Tropenland Wirt Zeitschrift fur die Landwirtschaft in den Tropen und Subtropen*. 1993; 94: 37-44.
  46. Pandey IP, Ahmed SF, Chhimwal S, Pandey S. Chemical composition and wound healing



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- activity of volatile oil of leaves of *Azadirachta indica* A. juss. *Advances in Pure and Applied Chemistry*. 2012; 62: 2167-0854.
47. Panda DN, Mishra A, Mishra SC, Rao AG. Efficacy of Esb3@in the treatment of experimental *Eimeria tenella* infection in broiler chicks. *Indian Veterinary Journal*. 1999; 76: 199-201.
  48. Pandey A, Pare P. A review: Antimicrobial activity of *Azadirachta indica* (Neem). *International Journal of Pharmacy & Life Sciences*. 2018, 9(3).
  49. Quraishi HA, Islam N, Iqbal A, Bhat SA, Ahmed J, Ashraf SS and Khan QA, (2018). Therapeutical and medicinal properties of Neem (*Azadirachta indica*) in context of Unani system of medicine: a review study. *Journal of Drug Delivery and Therapeutics*. 2018; 8(6-s), 394-399.
  50. Quiroz-Castañeda RE, Dantán-González E. Avian coccidiosis: future and present natural alternative. *BioMed Research International*. 2015; 11.
  51. Reddy YRR, Kumari CK, Lokanatha O, Mamatha S, Reddy CD. Antimicrobial activity of *Azadirachta indica* (Neem) leaf, bark and seed extracts. *International Journal of Research in Phytochemistry and Pharmacology*. 2013; 3(1): 1-4.
  52. Rahmani AH, Almatroudi A, Alrumaihi F, Khan AA. Pharmacological and therapeutic potential of Neem (*Azadirachta indica*). *Pharmacognosy Reviews*. 2018; 12(24): 250.
  53. Singh V, Chauhan D. Phytochemical evaluation of aqueous and ethanolic extract of Neem leaves (*Azadirachta indica*); *Indo American Journal of Pharmaceutical Research*. 2014; 4: 5943-5948.
  54. Saleque MA, Saha AA. Production and economic performance of small scale Sonali bird farming for meat production in Bangladesh. In *Proceedings of the Semian, 8th International Poultry Show and Seminar, Dhaka, World Poultry Science Association*. 2013: 20-24.
  55. Tewari AK, Maharana BR. Control of poultry coccidiosis: changing trends. *Journal of Parasitic Disease*. 2011; 35: 10-70.
  56. Tarhyel R, Hena SA, Tanimomo BK. Effect of age on organ weight and carcass characteristics of Japanese quail (*Coturnix japonica*). *Scientific Journal of Agriculture*. 2012; 1(1): 21-26.
  57. Teixeira M, Teixeira Filho WL, Lopes CWG. Coccidiosis in Japanese quails (*Coturnix japonica*): characterization of a naturally occurring infection in a commercial rearing farm. *Revista Brasileira de Ciência Avícola*. 2004; 6(2):129 - 134.
  58. Vongpakorn M, Adamu M, Boonkaewwan C, Gongruttananun N. Hematological, biochemical and histopathological changes caused by coccidiosis in chickens. *Kasetsart Journal (Nat. Sci.)*; 2013, 47 : 238 - 246.
  59. Xie MQ, Lu YM, Wen LN. Coccidiostatic and coccidiocidal test on coccidial oocysts of chickens. *Chinese Journal of Veterinary Medicine*. 1983; 9(10): 2- 5.
  60. Yusuf A, Awache I, Yakubu GM, Nosano M, Andefiki U, Maryfavour A. Phytochemical Properties and the in-vitro Antibacterial Activity of Neem (*Azadirachta indica*) Twigs Extract on Bacteria Isolated from Human Mouth. *Health Science Research*. 2017; 4(6): 68-71.